# Enhanced NK and CD8+ T cell proliferation, tumor cytotoxicity and reversal of T cell exhaustion with IGM-7354, an anti-PD-L1 IgM antibody and IL-15 cytokine fusion Thierry Giffon, Melanie Desbois, Poonam Yakkundi, Keerthana Sekar, Marigold Manlusoc, Rodnie Rosete, Daniel Machado, Susan Calhoun, Tasnim Kothambawala, Dean Ng,

### Background

Anti-PD-1/PD-L1 therapies are efficacious in certain cancer indications, but often patients relapse following a primary response. Therefore rational combinations are needed to enhance initial and durable responses of anti-PD-1/PD-L1 therapies. Immunostimulatory cytokine, IL-15 is an attractive combination partner to enhance antitumor NK and memory CD8<sup>+</sup> T cell expansion and survival.

We are developing IGM-7354, a high affinity, high avidity anti-PD-L1 pentameric IgM antibody with an IL-15R $\alpha$  chain and IL-15 fused to the joining (J) chain, designed to deliver IL-15 to PD-L1 expressing tumors to enhance anti-tumor immune responses.



Figure 1. A) Structure of IGM-7354. B) Natural presentation of IL-15 versus presentation of IL-15 by IGM-7354 to CD8 or NK cells. The presentation of IL-15 on the PD-L1 IgM can occur on both antigen presenting cells (APCs) and/or PD-L1-expressing tumor cells.

### IGM-7354 binds to human and cynomolgus monkey PD-L1 and IL-15 receptor β chain



Figure 2. Binding by ELISA to recombinant PD-L1 and IL-15 receptor chains. A) Species specificity was tested by ELISA with IGM-7354 and recombinant PD-L1 from human (hu), cynomolgus monkey (cy), rat (rt) and mouse (ms). B) Blockade of huPD-1/PD-L1 by IGM-7354 with PD-1/PD-L1 Blockade Bioassay. **C)** Binding of IGM-7354 to human IL-15 receptor α, β and γ chains. **D)** Binding of IGM-7354 to recombinant IL-15 receptor β chains from human, cynomolgus monkey, rat and mouse. Recombinant proteins were coated at 1 µg/mL and binding of IGM-7354 was measured using an anti-human kappa secondary antibody.

# IGM-7354 induces proliferation of NK and CD8+ T cells and phosphorylation of STAT5 in vitro



Figure 3. In vitro potency of IGM-7354 in PBMC cultures. A) Cell proliferation was determined by Ki67 staining of NK, CD8+ and CD4+ T cells after 96hr incubation with IGM-7354. B) Phosphorylation of STAT5 was measured by phosphoflow in NK, CD8+ and CD4+ T cells after 4hr incubation with IGM-7354. No Ki67 or pSTAT5 signals were observed with an anti-PD-L1 control IgM lacking the functional IL-15 fusion on the J chain (data not shown).

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cells (T<sub>ex</sub>), pan-T cells (CD3+) were isolated from PBMC donors and repeatedly stimulated using CD3/CD28 Dynabeads®. At the end of the final round of stimulation (day 6) Dynabeads® were removed and the T cells rested for a further 48 hours before inclusion in the MLR assay. **B)** T<sub>ex</sub> were phenotyped by flow cytometry to confirm expression levels of PD-1, TIM-3 and LAG-3 by CD8 T cells (on day 0 and day 8). Proliferation was assessed by staining for Ki67. Supernatant collected throughout the process was assessed for IL-2 by ELISA. IGM-7354 promoted increased secretion of IFNγ, and proliferation (Ki67) by exhausted T cells. One-way mixed lymphocyte reactions (MLRs) were used to assess reversal of T<sub>ex</sub> hypo-responsiveness in the presence of dose titrations of IGM-7354 and control molecules. In vitro generated monocytes-derived DCs (mo-DCs) were combined with T<sub>ex</sub> (1:10 ratio) to generate MLR pairs and cultured for 5 days. C) Following completion of the MLR, the supernatants were assessed for levels of IFNy (by ELISA). D) Additionally, the T cell populations were assessed for proliferative responses to the allogeneic stimulus (% Ki67 in CD8 populations). Data are representative of N=6 MLR pairs. Atezolizumab was purchased from Selleckchem.

Control MLRs were used to assess reversal of exhaustion: Control T cells (freshly isolated) + moDCs (Tcon+moDC), T exhausted + moDC + anti-PD-1 IgG (Tex+moDC+Nivo).

The T cell exhaustion studies were completed at Antibody Analytics Ltd., BioCity Scotland

experiment. MDA-MB-231-Luc cells (5000 cells) were incubated for 6 days in the presence of whole PBMCs at a 3:1 E:T ratio( E=CD8+NK). MDA-MB-231 cell killing was measured by addition of BioGlo (Promega) Shown is a representative experiment of dose dependent killing of the cancer cells. PD-L1 IgM has the same PD-L1 binding units as IGM-7354 but lacks the IL-15 on the joining chain. MDA-MB-231 were used in an in vivo tumor xenograft model (study # 1). MDA-MB-231 tumor cells (5x10<sup>6</sup>) were implanted s.c. in MHC-/- NSG mice. PBMCs (10<sup>7</sup>) were engrafted i.v. 10 days later. IGM-7354 or vehicle was dosed i.p. 2 days later at 5 or 20 mg/kg with the following regimen: Q2d x3, for 3 series of administration (9 doses total) with a week rest between administrations. **B)** Individual tumor size at Day 61 post-dosing. **C)** Individual tumor growth curves. \*\*\*\* p < 0.0001, \*\*\* p = 0.0003, ns non-significant (One-way ANOVA, with Tukey's multiple comparison test) In vivo MDA-MB-231 study # 2 with titration of IGM-7354 dosed at 1, 3 and 10 mg/kg. D) Average tumor

Figure 6. A) PD-L1 expressing MDA-MB-231-Luc cells were used as targets in an in vitro killing

growth curves. E) Absolute counts of circulating NK and CD8 T cells, and proliferation of NK and CD8 T cells determined by Ki67 staining on whole blood samples at Day17.



10 suggests synergy.

**RPMI-8226 were used in an** *in vivo* **tumor xenograft model.** RPMI-8226 tumor cells (1x10<sup>7</sup>) in PBS/Matrigel (1:1) were implanted s.c. in CB17/SCID mice. 14 days later, mice were randomized and IGM-7354 or vehicle was dosed i.p. at 20 mg/kg with the following regimen: Q2d x3, for 3 series of administration (9 doses total) with a week rest between administrations. Daratumumab was dosed i.p. biweekly for 5 weeks. C) Average tumor growth curves. D) Individual tumor size at Day 28 post-randomization. \* p = 0.031, \*\* p = 0.0091, ns non-significant (One-way ANOVA, with Tukey's multiple comparison test).

- IND filling planned for 2022

Days post-tumor implantation

# IGM-7354 demonstrates additivity and potential synergy in combination with ADCC-mediating antibodies

(E=NK) with the ADCC-enabling antibodies added for the last 24 hours. Target cell killing was measured by addition of BioGlo (Promega). A) Shown are representative experiments of dose dependent killing of the cancer cells. **B)** Bliss synergy scores were calculated in SynergyFinder and visualized in 3D surface plots. Bliss score >

## **Summary**

### IGM-7354, an anti-PD-L1 –IL-15 fusion IgM antibody:

Binds equivalently to human and cynomolgus PD-L1 but not rodent.

• Binds equivalently to human and cynomolgus IL-15 receptor  $\beta$  chain through its IL-15 fusion on the J chain. Binding to rodent receptors was 10-15-fold weaker.

• Provides a potent IL-15-dependent proliferation signal to primary human NK and CD8+ T cells in vitro, as measured by Ki-67 and STAT5 phosphorylation.

• Enhances in vitro killing of PD-L1 positive MDA-MB-231 cells by human PBMCs compared to anti-PD-L1 IgM lacking the IL-15 fusion.

• Demonstrates dose-dependent responses in a human MDA-MB-231 xenograft mouse tumor model, with some treated animals having complete tumor regressions.

• Shows additivity and potential synergy in combination with antibodies with ADCC mechanism of action such as Cetuximab, Daratumumab, and Trastuzumab in vitro and in vivo.

· Reverses T cell exhaustion more potently than anti-PD-L1 antibodies or a nontargeted IL-15 fusion molecule alone in an in vitro modified MLR model.

• Pharmacodynamic studies in cynomolgus monkeys shows potent proliferation of circulating NK, CD8+  $T_{FM}$  and  $\gamma\delta$  T cells over the course of the treatment.

• IGM-7354 may enhance tumor localization of immunostimulatory cytokine IL-15 through the high affinity and high avidity binding to PD-L1 to improve antitumor responses and minimize toxicity.