A Phase 1 Dose Escalation Study of IGM-2323, A Novel Anti-CD20 x Anti-CD3 IgM T Cell Engager (TCE) in Patients with Advanced B-Cell Malignancies

*Elizabeth Budde, MD PhD*¹, *Ajay K. Gopal, MD*², *Won Seog Kim, MD PhD*³, *Ian Flinn, MD PhD*⁴, *Chan Y. Cheah, MBBS*⁵, *Loretta Nastoupil, MD*⁶, *Matthew Matasar, MD*⁷, *Catherine Diefenbach, MD*⁸, *Gareth P. Gregory, MBBS PhD*⁹, *Ibrahim Qazi, PharmD*¹⁰, *Ching–Fai Pang, PhD*¹¹, *Maya Leabman, PhD*¹⁰, *Genevive Hernandez, PhD*¹⁰, *Iris Sison*¹⁰, *Bruce Keyt, PhD*¹⁰, *Daniel Chen, MD PhD*¹⁰, *and Philippe Armand, MD PhD*¹²

¹T Cell Therapeutics Research Laboratory, Department of Hematology and Hematopoietic Cell Transplantation, City of Hope National Medical Center, Duarte, CA

²University of Washington/Fred Hutchinson Cancer Research Center/Seattle Cancer Care Alliance, Seattle, WA

³Samsung Medical Center, Seoul, South Korea

⁴Sarah Cannon Research Institute/Tennessee Oncology, Nashville, TN ⁵Linear Clinical Research and Sir Charles Gairdner Hospital, Perth, Australia

⁶The University of Texas MD Anderson Cancer Center, Houston, TX

⁷Memorial Sloan Kettering Cancer Center, New York, NY

⁸Perlmutter Cancer Center at NYU Langone Health, New York, NY

⁹School of Clinical Sciences at Monash Health, Monash University, Melbourne, Australia

¹⁰IGM Biosciences, Mountain View, CA

¹¹Phi Consulting Group, Bellevue, WA

¹²Department of Medical Oncology, Dana Farber Cancer Institute, Boston, MA

Introduction:

IGM-2323 is the first engineered high-affinity, high-avidity bispecific IgM monoclonal antibody TCE to be tested in the clinic. It has 10 binding domains for CD20 and a single binding domain for CD3 and is designed to bind irreversibly to CD20-high and low-expressing cells with more physiologic stimulation to T cells, which may mitigate cytokine release syndrome (CRS)-related toxicity and broaden the therapeutic window. IGM-2323 may act by multiple mechanisms: T-cell dependent cytotoxicity, complement dependent cytotoxicity, and enhanced immune modulation via IFNγ-dominant cytokine

stimulation. This phase 1 study is exploring the safety and activity of IGM-2323 using a dose titration schedule intended to optimize repeatable immune stimulation while minimizing toxicity.

<u>Methods</u>:

This first-in-human Phase 1 study is a global, multicenter, open-label, dose escalation evaluating safety, tolerability, PK, and preliminary efficacy (NCT04082936). Adults with relapsed or refractory CD20⁺ B-cell NHL with ≥ 2 prior systemic therapies, adequate organ function, and ECOG 0-1 are eligible. IGM-2323 is given IV on Days 1, 8, and 15 of 21-day cycles until disease progression or unacceptable toxicity. Treatment can continue beyond progression if the patient (pt) has benefitted from treatment and intra-patient dose escalation is allowed. This study also utilizes a dose titration scheme where a starting dose is given on Day 1, then higher subsequent doses are given up to a plateau dose. If a pt has symptoms of CRS, the same dose may be repeated, and a higher dose is not given until the dose is well tolerated.

<u>Results</u>:

As of April 30, 2021, 29 pts have been enrolled: 12 at 5 fixed dose levels (0.5, 2.5, 10, 30, 100 mg) and 17 at 5 dose titration levels (50/100, 50/200, 50/300, 50/600, 50/1000 mg)*.

NHL subtypes include: 13 follicular (FL), 11 diffuse large B-cell (DLBCL), 3 mantle cell (MCL), and 2 marginal zone (MZL). Median age is 66 (range, 36–84) and median prior therapies is 3 (range, 2–7). 2 pts had prior autologous transplant and 7 had prior CAR-T.

All 29 pts received at least one dose and are safety evaluable. There were no DLTs and no neurotoxicity AEs. No pts discontinued due to AE. There were 3 drug-related SAEs (1 each Gr1/2/3 CRS by ASTCT). 16 pts discontinued treatment: 2 pt/investigator decision and 14 PD. The 13 pts still on treatment have received a median of 9 doses (range, 1–42).

At fixed dose levels, 5 of 12 pts had a CRS event, primarily grade 1 (fever). Only 1 of 17 titration pts had CRS (Gr3 in a pt with prior experimental CAR-T and baseline circulating B-cells). The most common drug-related AEs in titration pts were hypophosphatemia (29%), IRR (29%; the distinction between IRR and CRS was determined by the investigator), and nausea (24%).

23 pts were evaluable for efficacy; there were 8 responses (5 CR, 3 PR). 5 responses were in FL/MZL and 3 in DLBCL/MCL. Response kinetics varied, with some CRs at the first scan (Week 6) and others after biopsy-confirmed pseudo-progression or prolonged SD (up to 69 weeks). 7 of 8 responses were ongoing as of the data cut. 5 of 11 evaluable patients in dose titration cohorts responded (3 CR, 2 PR), with a median time to response of 6 weeks. In the titration cohort with the longest follow-up (50/100), there were 2 CR and 1 PR in 4 evaluable pts.

Repeatable IFN γ stimulation is detectable in nearly all pts with measurable cytokine elevations, while polycytokine response was observed in CRS cases (Table). Preliminary IHC data in matched baseline and on-treatment biopsies (n=3) shows decrease of CD20⁺ tumor cells and increase of CD3⁺ T cells in tumor tissue upon treatment, regardless of clinical response.

Conclusions:

Preliminary results from this first-in-human study of IGM-2323 show an excellent safety and tolerability profile at up to 1000 mg, with reduced CRS when IGM-2323 was given using a dose titration scheme. Response patterns included rapid responses, deepening responses and pseudo-progression. IGM-2323 also has evidence of an IFNγ-dominant repeatable T-cell activation and preservation of T cell function over time. The MTD of IGM-2323 has not been reached. Clinical activity was observed across multiple histologies. Updated safety, PK, biomarker, and efficacy data, including complete dose escalation data, will be presented at the meeting.

* starting and plateau dose referenced

Peak Cytokine Levels Post-infusion (Cycle 1)

Cytokine Median (IQR) pg/mL	No CRS (n=22)	CRS (<i>n=6</i>)
IFN-γ	37 (101)	756 (3841)
IL-6	6 (9)	346 (2550)
TNF-α	12 (15)	68 (192)

Cytokine levels in frozen plasma were assessed on a batched basis at a central lab. The table shows median values and interquartile range (in parenthesis) of the highest concentrations obtained in each patient during the sampling period: 30 min to 72 hours for Infusion 1 and end of infusion to 24 hours for Infusions 2 and 3.